

2019-03

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<http://hdl.handle.net/10026.1/14357>

10.1111/ene.13837

European Journal of Neurology

Wiley

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Neurological effects of glucocerebrosidase gene mutations

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Gaucher, GBA, glucocerebrosidase, neuronopathic, Parkinson

Received 16 May 2018
Accepted 9 October 2018*European Journal of Neurology* 2019, **26**: 388–393,
e26–e29

doi:10.1111/ene.13837

The association between Gaucher disease (GD) and Parkinson disease (PD) has been described for almost two decades. In the biallelic state (homozygous or compound heterozygous) mutations in the glucocerebrosidase gene (GBA) may cause GD, in which glucosylceramide, the sphingolipid substrate of the glucocerebrosidase enzyme (GCase), accumulates in visceral organs leading to a number of clinical phenotypes. In the biallelic or heterozygous state, GBA mutations increase the risk for PD. Mutations of the GBA allele are the most significant genetic risk factor for idiopathic PD, found in 5%–20% of idiopathic PD cases depending on ethnicity. The neurological consequences of GBA mutations are reviewed and the proposition that GBA mutations result in a disparate but connected range of clinically and pathologically related neurological features is discussed. The literature relating to the clinical, biochemical and genetic basis of GBA PD, type 1 GD and neuronopathic GD is considered highlighting commonalities and distinctions between them. The evidence for a unifying disease mechanism is considered.

Introduction

Gaucher disease (GD) is an autosomal recessive disorder caused by mutations of the glucocerebrosidase gene (GBA). The GBA encodes the lysosomal hydrolase glucocerebrosidase (GCase), which under acidic conditions will hydrolyse the sphingolipid waste product glucosylceramide into ceramide. Mutations cause a reduction or complete loss of GCase activity which (in the case of some biallelic subjects) leads to the accumulation of glucosylceramide (and its deacylated derivative glucosylsphingosine) in visceral organs, causing a variety of clinical phenotypes. These include thrombocytopaenia, anaemia, hepatosplenomegaly and osteopaenia/osteonecrosis [1,2]. A subset of these patients will also develop central nervous system (CNS) features [1].

Gaucher disease is broadly categorized into three subtypes. Type 1 GD (GD1) is diagnosed in the

absence of central neurological features, although peripheral nervous system involvement in the form of a symmetrical polyneuropathy does occur [3]. Type 2 GD (GD2) is characterized by rapid neurological deterioration and is ultimately fatal whilst in type 3 GD (GD3) there are slower progressive neurological features. In practice, the distinction between GD2 and GD3 are often blurred and it has been suggested that the exact neurological phenotype is more likely to be a spectrum disorder [4]. For the purpose of this review, types 2 and 3 will be referred to collectively as neuronopathic Gaucher disease (nGD).

The neurological phenotype of nGD

There is a substantial variation in the neurological features associated with nGD. A supranuclear gaze palsy is the most common clinical presentation; however, this may be because it is an easily identified clinical feature which allows the diagnosis to be readily made [1,4–6]. This advances the question whether in fact the definition of nGD is an arbitrary waypoint in

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a spectrum of neurological features present across nearly all Gaucher patients [7].

Otherwise cognition is the second most common feature of nGD [5] in combination with a variety of other features such as tremor, myoclonus, seizures, gait disorders, bulbar dysfunction, stridor, muscle weakness [1]. There appears to be some regional variation between this phenotype [3,8–10], which may or may not be explained by variation in GBA mutations present [11]. Equally, attempts to show mutation-specific phenotypes have been unsuccessful [4,7].

Central nervous system features in GD1

A minority of GD1 patients are known to develop peripheral neuropathy [3]; however, the prevailing view has been that GD1 patients do not develop central nervous system (CNS) involvement. In recent years, there have been grounds to consider the need for revision of this view. The discovery that GD1 patients have a lifetime risk of developing Parkinson disease (PD) of approximately 10%–30% compared to approximately 1%–2% in the general population indicates that CNS dysfunction as a consequence of a GBA mutation can be lifelong. A study of the cognitive profile of Gaucher patients found that, compared to age-matched controls, there was delayed recall of verbal and non-verbal information along with reduced attention [8].

Our own prospective data have shown general cognitive impairment along with impaired olfaction amongst not only GD1 patients but also ‘asymptomatic’ heterozygous GBA carriers [12,13]. Moreover, deterioration of cognition, olfaction and depression was more marked amongst both these groups compared to controls. It is notable that the distribution of these features correlates with the sequence of prodromal symptoms that characterize the preclinical phase of PD.

Glucocerebrosidase gene mutations and PD

Although a family history of idiopathic PD results in approximately a fourfold increase in lifetime risk [14], it is not ordinarily passed on in a Mendelian manner. The recognition that GBA mutations are an important risk factor for PD has proven to be an important stimulus for additional insights into the pathogenesis of PD. Unlike other genetic forms of PD, GBA mutations display incomplete lifetime penetrance of around 10%–30% [3,4,7]. The GBA mutation minor allele frequency is 1% in the general population [12,13], 5% in the Ashkenazi population [3,15,16], between 5% and 10% in idiopathic PD [8] and between 10%–30%

in Ashkenazi Jews with PD [3,15,16]. The precise frequencies in the respective populations depend upon whether the whole exome has been sequenced, whether targeted microarray genotyping of specific (common) mutations is carried out and upon the inclusion or not of mutations such as E326K which is not associated with Gaucher disease [12,13] but is associated with an increased risk of PD [17].

The neurological phenotype of GBA PD

Although there is some debate about the GBA PD phenotype, it appears to be associated with a more pronounced cognitive deficit. This is supported by data which implicate GBA in dementia with Lewy bodies, a form of dementia with associated parkinsonism and other differentiating features such as hallucinations [18]. Equally there are some data to suggest that there is an enhanced neuropsychiatric phenotype. Neuroanatomically, this has been considered to suggest a more ubiquitous spread of Lewy pathology across cortical brain regions, in contrast to PD which predominantly affects the basal ganglia, although increasingly it is accepted that Lewy deposits affect a variety of brain regions, especially in late idiopathic PD [19,20]. That said, asymmetrical reductions in dopamine uptake have been demonstrated amongst GBA PD subjects within the basal ganglia [21].

The clinical course of GBA PD has conclusively been shown to be more aggressive, with a younger age of onset compared to idiopathic PD and a lower median survival time from diagnosis [22]. Some data indicate that classical features of PD including asymmetrical onset, bradykinesia, postural instability and rigidity are less pronounced in GBA PD subjects [23]. To date, no genetic association has been found with other synucleinopathies such as multisystem atrophy [3].

Mechanistic explanations for nGD and GBA PD

There is no consensus on the underlying mode of action of neurodegeneration in nGD and GBA PD. It is beyond the scope of this paper to review these data in detail; however, broadly they can be categorized into those which postulate a loss of function and those which postulate a gain of function. The argument of a loss of function centres primarily on the finding of reduced GCase catalytic activity in both GD (blood) and GBA PD [blood and cerebrospinal fluid (CSF)] [11,24,25]. However, there are major inconsistencies in this argument. In particular, there is a relatively poor correlation between GCase activity levels and disease risk in PD and GD. For instance,

one study showed that GCase activity from peripheral blood spots categorized by mutation did not correlate with the severity of those individual mutations in terms of PD. L444P PD cases for instance appear to have higher GCase activity than those with N370S, even though L444P mutations convey between two and three times the risk of PD compared to N370S [11,26,27]. Equally, heterozygous carriers of the 84GG mutation, which is a frameshift mutation and would be expected to be null in terms of GCase activity, had comparable enzyme activity with other missense GBA mutations [28,29]. Although in GD residual GCase activity has an influence, it does not in isolation predict disease prognosis [28]. A number of mechanisms have been proposed by which loss of function could lead to GD and PD. Predominantly, these are accumulation of enzyme substrate (GD and PD) and failure of autophagic pathways leading to reduced disposal of alpha synuclein [30].

It may be that this disparity is a reflection of the limitations of the GCase activity assay. In leucocytes GCase is typically normalized to the protein concentration of the lysate produced and in CSF there is no normalization to protein concentration at all [25,31]. Differences in the expression levels of the mutant and wild-type GCase protein (from the remaining non-mutated allele) will substantially influence what is effectively an assay that measures the rate of substrate catalysis, yet to date there is no mechanism to normalize for GCase protein expression.

Conversely, some argue that GBA induces a toxic gain of function. Perhaps the most convincing of these explanations comes from the work of Mia Horowitz who suggests that sequestration of mutant GBA within the endoplasmic reticulum, due to a failure of the normal process of post-translational folding, leads to a variable degree of unfolded protein response which correlates with the pathogenicity of the mutation [32].

There may of course be the potential for both loss of function and a toxic gain of function to be coexistent or perhaps more plausibly a product of one another. For instance, any structural alteration to the GCase protein may give rise to gain in function but may incidentally reduce enzyme activity. Conversely, a loss of enzyme activity may result downstream in an added gain of function as a result of disruption of the balance of substrate/product.

Neuropathology of GD and PD

Parkinson disease is pathologically well defined by the degeneration of the substantia nigra pars compacta and the presence of Lewy bodies. These are protein inclusions predominantly comprising alpha synuclein,

a ubiquitously expressed cytosolic protein of unknown physiological function that appears to be central to the pathological processes that contribute to PD [33,34]. Its key importance is confirmed by the finding that missense mutations of the alpha synuclein gene (SNCA) cause autosomal dominant familial PD [24,25] whilst polymorphisms in SNCA cause a marginal protective/causative effect in terms of developing the PD phenotype [11]. Interestingly missense mutations in the Rep1 promotor region [11,26,27] and duplications/triplication of the whole SNCA gene [28,29] are also implicated or causative of PD, with the age of onset being proportional to the expression levels of wild-type alpha synuclein protein. Lewy pathology is present in around 20% of subjects of over 80 years without any signs of parkinsonism, indicating that alpha synuclein accumulates with age, a process that reflects the age-related incidence of PD [28].

A recent concept in the pathophysiology of PD is that alpha synuclein may be transmitted in a prion-like fashion [30]. Briefly, alpha synuclein is a monomeric protein in the native state; however, under a variety of intracellular conditions (such as proximity to a lipid membrane [35] or oxidative stress [36]) it converts to a beta sheet rich fibrillar/oligomeric and then an aggregated form [37]. It is this aggregated phosphorylated form that is the major component of Lewy bodies [38]. Postmortem examinations of PD patients who received stem cell grafts as part of a clinical trial showed that within a decade transplanted neuronal tissue also contained Lewy pathology [39]. It appears that aggregated alpha synuclein can 'seed' monomeric forms to adopt these fibrillar/oligomeric/aggregated configurations, which in turn can propagate Lewy pathology to adjacent cells [40–43].

Some studies have demonstrated Lewy pathology in the enteric nervous system [44–46] and have suggested that it may propagate along peripheral nerves (most prominently the vagus nerve), through the midbrain and then subsequently into the cortex and neocortex, with PD severity correlating with the extent of the sequential spread of this pathology [47,48]. Concurrently, a series of prodromal features of PD sequentially occur in a manner which is postulated to correlate with the pathological spread [44,49,50].

Neuronopathic Gaucher disease has been less well characterized. There are only a few postmortem studies of nGD patients, a substantial portion of which were performed two to three decades ago [51–54]. There have also been a number of mouse models of nGD, although most of these employ artificial disruption of the GBA gene to produce a broadly comparable phenotype and are of questionable clinical

relevance [55,56]. A common feature appears to be microgliosis and subsequent astrogliosis and neuronal loss, particularly in hippocampal regions [51,55], along with substrate accumulation [52,54,55,57,58]. However, there appears to be significant variation in the severity of neuropathology in nGD cases, which is broadly proportional to the degree of CNS involvement [51]. A postmortem study of three GD1 patients (all N370S homozygous or N370s compound heterozygous) without clinical neurological involvement revealed microgliosis and astrogliosis in cortical and hippocampal regions [51]. This glial activation mimics the distribution of hippocampal pathology in nGD patients, with the distinction that amongst nGD subjects there is accompanying neuronal loss [51].

Recent data suggest that the deacylated derivative of glucosylceramide, glucosylsphingosine, may be the pathogenic molecule in nGD. Most pertinently glucosylsphingosine appears to be raised far more specifically in (plasma) GD subjects than other biomarkers including glucosylceramide [33,34]. Unfortunately, only a few more recent studies have attempted to quantify neuronal levels of the lipid; however, it appears to be present in human nGD [54] and transgenic mouse brains [55].

Cell biology of GD and GBA PD

Glucocerebrosidase gene mutations may cause aberrant post-translational folding of the enzyme. This prevents trafficking of the enzyme from the endoplasmic reticulum to the lysosome and causes the GCase to become sequestered within the endoplasmic reticulum [6,59,60]. The consequences of this are twofold. On the one hand, insufficient GCase reaches the lysosome to allow sufficient catalysis of glucosylceramide to ceramide. The sequestered GCase also gives rise to an unfolded protein response which in turn results in oxidative stress in the cytosol [6,59–61].

In vitro studies have highlighted a bidirectional inverse correlation between GCase activity and alpha synuclein [62,63]. Concurrently, biophysical studies have shown a direct interaction with alpha synuclein which is lessened for GBA mutation carrying forms of the enzyme [64,65]. These findings are within the broader context of an established literature implicating diminished mechanisms of cellular autophagy in idiopathic PD [66–68]. Interestingly, there appears to be an age-dependent reduction in GCase activity in both mice and human brain [69,70]. Moreover, GCase activity levels are reduced compared to controls in the brains of non-GBA idiopathic PD subjects [71]; however, studies looking at substrate accumulation are somewhat varied. Some show no increase in

glucosylceramide or glucosylsphingosine levels in PD or aged controls [72,73] but another found an increase in the latter [69]. Mouse data have also shown age-dependent upregulation of glucosylsphingosine and two studies found that ceramide levels (the product of substrate breakdown) are reduced in the brains of GBA PD subjects [74,75]. Thus, although there is evidence to support a role for GCase in the proteolysis of alpha synuclein, with GBA mutations causing reduced GCase activity and in turn reduced turnover of cytosolic alpha synuclein, the precise mechanism underlying this remains unknown.

Parallels between nGD and GBA PD

Variable penetrance of nGD and GBA PD

A particularly pertinent aspect of the natural history of nGD and GBA PD is the variable penetrance of the phenotype in both conditions (i.e. many with compatible genotypes will not develop PD, GD1 or nGD). The mutation/phenotype correlation is not absolute, although similar patterns exist across the two conditions. For instance severe GBA mutations, which by definition are associated with nGD [76], increase the risk of PD by around four times compared to mild ones [26]. Equally, specific mutations which are known to be highly pathogenic in terms of nGD (such as L444P) [77] are also amongst the most pathogenic in terms of PD risk [78]. Conversely those which have a questionable effect on the GD phenotype [24,25], such as E326K and T369M, deliver only a marginal increase in PD risk [11].

Stratification of disease risk by categorization of mutations has aroused significant interest in the context of PD [11,26,27]. The severe/mild dichotomy has been applied prospectively to PD cohorts, and it has been shown to have a major impact on age at onset of PD and median time to dementia [28,29]. It may allow more effective targeting of future disease modifying treatments for PD, increase the power of clinical trials of such compounds by allowing selection of subjects who are likely to have a more rapid decline [28]. Moreover, the fact that the same mutations determine the severity of neurological features in both GD and PD suggests that a common mode of action is shared by the two conditions.

Neuroinflammation in GBA PD and nGD

There is evidence for immune dysregulation associated with, and putatively causative of, a number of the major clinical phenotypes of GD [79–82]. Additionally, there is substantial evidence of an increased risk

of monoclonal gammopathy, multiple myeloma as well as lymphoid malignancies within GD patients [83,84]. GD patients have been shown to have increased serum concentrations of interleukin 6 and 10 [79] whilst GBA knockout mice display upregulation of a variety of immune markers and increased levels of a variety of inflammatory cytokines in the plasma [85,86]. This has been hypothesized to be a result of accumulated glucosylceramide within the mononuclear phagocytic system, although the precise mechanism is still unclear [85].

Immune activation has also been highlighted as a key factor in the pathogenesis of PD [87]. Substantial *in vitro* data have directly linked alpha synuclein to microglial activation, with a number of mechanisms postulated [88–94]. This correlates with *in vivo* clinical imaging studies [95], postmortem evidence [96,97], serum titres of cytokines [98,99] and autoantibodies to components of the synuclein aggregation pathway [100]. Interestingly chronic use of non-steroidal anti-inflammatory drugs has been shown to exhibit a mildly protective effect against PD onset [14]. One study has also shown enhanced activation of a number of peripheral cytokines within GBA PD patients [101].

Disease modifying therapy in GD, nGD and GBA PD

Enzyme replacement therapy

The advent of recombinant enzyme replacement therapy (ERT), given intravenously, usually on an approximately monthly basis, has transformed the prognosis for those with GD [2]; the majority of GD1 patients can now live practically normal lives. The exception to this is nGD, which is at present unresponsive to ERT as the 596 kDa protein is unable to cross the blood–brain barrier [2,102]. There are various strategies under consideration which aim to temporarily permeabilize the blood–brain barrier to allow recombinant ERT into the brain [103]. These include use of a lentiviral vector to chaperone the protein [104,105]. Intrathecal administration of ERT has been used with some success in paediatric Hurler's syndrome (mucopolysaccharidosis type 1) [106]. Moreover, a trial of intrathecal ERT in Hunter's disease (mucopolysaccharidosis type 2) is also under way [107]. That said, there are a number of pitfalls which must be overcome. The major challenge is ensuring the ERT is able to breach the cell membrane and reach the intracellular compartment. There is also the very real danger of an immune mediated CNS inflammatory reaction following such administration [106].

Substrate reduction therapy

Other strategies to overcome the issue of CNS penetrance include substrate reduction therapy. Here inhibition of upstream enzymes in the metabolic pathway such as glucosylceramide synthase actively reduce substrate production. Miglustat is one such compound. Mouse models have shown, using microglial inflammation as a surrogate marker of CNS penetrance, that miglustat crosses the blood–brain barrier [108] and there is some anecdotal evidence of clinical efficacy in GD3. However, a small randomized controlled trial failed to prove this, although this may be on account of the poor choice of primary outcome, namely an improvement in supranuclear gaze palsy [109,110]. Equally its use was associated with an apparently increased incidence of peripheral neuropathy [110,111].

In vitro studies have suggested that, in cell lines expressing autosomal dominant alpha synuclein mutations, these therapies reduce levels of the synuclein protein [17], which has led to a phase II clinical trial of the glucosylceramide synthase inhibitor venglustat as a putative neuroprotective in GBA PD (clinicaltrials.gov, NCT02906020). Whilst intriguing, substrate accumulation has not consistently been demonstrated in PD brains. Nevertheless, it may be that minute undetectable alterations in substrate levels interact with alpha synuclein in other neurotoxic ways, such as for instance by altering the aggregation dynamics of the protein [112].

Small molecular chaperones

Another therapeutic strategy is the use of small molecular chaperones which are also able to enter the CNS. These compounds appear to modify the tertiary and quaternary structure of GCase, facilitating post-translational folding and transport of GCase to the lysosome [113]. At the lysosome the change in pH causes the chaperone to dissociate from GCase allowing effective catalysis to resume. A recent phase II trial of ambroxol, a repurposed cough linctus which also acts as a small molecular chaperone to GCase, showed good CSF penetrance and a reduction of glucosylceramide levels in the CSF [114]. Similarly our own group is currently conducting a phase II trial of ambroxol in PD patients (NCT02941822).

Gene therapy

Interest in gene therapy targeting the GBA gene up to this point has been predominantly in the context of GD [18]. Of particular interest has been the realistic

prospect of such therapy crossing the blood–brain barrier and hence being a viable treatment for neurological GBA related disease. Using invariant natural T killer cells, it was shown in 2011 that Gaucher fibroblasts could be transfected using a high-titre, amphotropic retroviral vector in which human GBA1 was driven by the mutant polyoma virus enhancer/herpesvirus thymidine kinase gene promoter [19,20]. Unfortunately the clinical protocol that resulted achieved a transfection efficiency of only 1%–10% which was insufficient to cause a clinically significant improvement in substrate levels [21]. Using a murine GBA knockout model, an intravenously injected lentiviral vector was shown to be able to deliver improved GCase activity, reduced substrate accumulation and improvements in a number of clinical outcomes [22]; however, whether such outcomes can be replicated in human subjects is uncertain.

In the context of PD, moreover, it is unclear whether affected brain regions (e.g. the substantia nigra and striatum) could be reached by such vector technologies. However, systemic administration to transgenic synuclein overexpressing mice of AAV9-PHP.B (an AAV which readily crosses the blood–brain barrier) resulted in complete clearance of alpha synuclein throughout the brain [22]. The approach has great promise as a means of readily crossing the blood–brain barrier; however, finding stable and safe means of efficient transfection remains the principal obstacle to its successful clinical application [23].

Concluding remarks

In this paper, it is suggested that in terms of CNS involvement the traditional dichotomization of GD1 and nGD patients may require revision and that a continuum of neurodegeneration may exist across biallelic and heterozygous carriers of GBA mutations. The evidence for this is mixed. The same (severe) mutations are associated with a worse phenotype in both diseases and the penetrance of this phenotype is

similarly variable. Neuropathologically, the presence of substrate deposits in the brains of nGD patients and their low or absent levels in heterozygous GBA PD cases imply no commonality of pathology across the two diseases. However, postmortem studies of nGD and GD1 patients have identified a pattern of microgliosis and then astrogliosis (with varying severity) which is anatomically very similar to that found in dementia with Lewy bodies. In this spectrum GD2 and then GD3 patients represent the more severe neurological phenotypes whilst GD1 patients and heterozygous GBA carriers display variable degrees of mild or subclinical neurological features. PD may simply be another neurological phenotype caused by GBA mutations.

No consensus exists regarding the pathogenic mechanism of nGD or GBA PD; however, a variety of promising disease modifying strategies, targeting a variety of proposed mechanisms, are currently in development or under evaluation. The authors believe that in the near future these approaches may elicit a tangible disease modifying effect in both diseases.

Acknowledgements

Funding was provided by the Cure Parkinson's Disease Trust (ref: AiM PD), the Leonard Wolfson Experimental Neurology Centre (PR/ylr/18575), the Medical Research Council (UK) (ref: MR/M006646/1), the UK Dementia Research Institute, the Kattan Trust (ref: Charity 285) and the Joint Programme of Neurodegenerative Disease Research (JPND) (ref: MR/N028651/1). AHVS is supported by the UCLH Biomedical Research Centre grant (ref: RCF73TS20145980) from the National Institute of Health Research (NIHR). SM is an NIHR funded academic clinical lecturer.

Disclosure of conflicts of interest

The authors declare no financial or other conflicts of interest.

References

- Grabowski GA, Zimran A, Ida H. Gaucher disease types 1 and 3: phenotypic characterization of large populations from the ICGG Gaucher Registry. *Am J Hematol* 2015; **90**: S12–S18.
- Grabowski GA. Phenotype, diagnosis, and treatment of Gaucher's disease. *Lancet* 2008; **372**: 1263–1271.
- Biegstraaten M, Mengel E, Marodi L, *et al.* Peripheral neuropathy in adult type 1 Gaucher disease: a 2-year prospective observational study. *Brain* 2010; **133**: 2909–2919.
- Goker-Alpan O, Schiffmann R, Park JK, Stubblefield BK, Tayebi N, Sidransky E. Phenotypic continuum in neuronopathic Gaucher disease: an intermediate phenotype between type 2 and type 3. *J Pediatr* 2003; **143**: 273–276.
- Abdelwahab M, Blankenship D, Schiffmann R. Long-term follow up and sudden unexpected death in Gaucher disease type 3 in Egypt. *Mol Genet Metab* 2016; **117**: S14.
- Tylki-Szymańska A, Keddache M, Grabowski GA. Characterization of neuronopathic Gaucher disease among ethnic Poles. *Genet Med* 2006; **8**: 8–15.
- Goker-Alpan O. Divergent phenotypes in Gaucher disease implicate the role of modifiers. *J Med Genet* 2005; **42**: e37.
- Biegstraaten M, Wesnes KA, Luzy C, *et al.* The cognitive profile of type 1 Gaucher disease patients. *J Inher Metab Dis* 2012; **35**: 1093–1099.
- Tajima A, Yokoi T, Ariga M, Ito T, Kaneshiro E, Eto Y, *et al.* Clinical and genetic study of Japanese patients with type 3 Gaucher disease. *Mol Genet Metab* 2009; **97**: 272–277.
- Lee J-Y, Lee BH, Kim G-H, *et al.* Clinical and genetic characteristics of Gaucher disease according to phenotypic subgroups. *Korean J Pediatr* 2012; **55**: 48–53.
- Zhang Y, Shu L, Sun Q, *et al.* Integrated genetic analysis of racial differences of common GBA variants in Parkinson's disease: a meta-analysis. *Front Mol Neurosci* 2018; **11**: 43.
- Beavan M, McNeill A, Proukakis C, Hughes DA, Mehta A, Schapira AHV. Evolution of prodromal clinical markers of Parkinson disease in a GBA mutation-positive cohort. *JAMA Neurol* 2014; **72**: 201–208.
- McNeill A, Duran R, Proukakis C, *et al.* Hyposmia and cognitive impairment in Gaucher disease patients and carriers. *Mov Disord* 2012; **27**: 526–532.
- Noyce AJ, Bestwick JP, Silveira-Moriyama L, *et al.* Meta-analysis of early nonmotor features and risk factors for Parkinson disease. *Ann Neurol* 2012; **72**: 893–901.
- Clark LN, Ross BM, Wang Y, *et al.* Mutations in the glucocerebrosidase gene are associated with early-onset Parkinson disease. *Neurology* 2007; **69**: 1270–1277.
- Gan-Or Z, Bar-Shira A, Mirelman A, *et al.* LRRK2 and GBA mutations differentially affect the initial presentation of Parkinson disease. *Neurogenetics* 2010; **11**: 121–125.
- Sardi SP, Viel C, Clarke J, *et al.* Glucosylceramide synthase inhibition alleviates aberrations in synucleinopathy models. *Proc Natl Acad Sci* 2017; **114**: 2699–2704.
- Kohn DB, Nolta JA, Weinthal J, *et al.* Toward gene therapy for Gaucher disease. *Hum Gene Ther* 1991; **2**: 101–105.
- Brennan PJ, Tatituri RVV, Brigl M, *et al.* Invariant natural killer T cells recognize lipid self antigen induced by microbial danger signals. *Nat Immunol* 2011; **12**: 1202–1211.
- Fink JK, Correll PH, Perry LK, Brady RO, Karlsson S. Correction of glucocerebrosidase deficiency after retroviral-mediated gene transfer into hematopoietic progenitor cells from patients with Gaucher disease. *Proc Natl Acad Sci USA* 1990; **87**: 2334–2338.
- Dunbar CE, Kohn DB, Schiffmann R, *et al.* Retroviral transfer of the glucocerebrosidase gene into CD34+ cells from patients with Gaucher disease: *in vivo* detection of transduced cells without myeloablation. *Hum Gene Ther* 2004; **9**: 2629–2640.
- Dahl M, Doyle A, Olsson K, *et al.* Lentiviral gene therapy using cellular promoters cures type 1 Gaucher disease in mice. *Mol Ther* 2015; **23**: 835–844.
- Cucchiari M. Human gene therapy: novel approaches to improve the current gene delivery systems. *Discov Med* 2016; **21**: 495–506.
- Walker JM, Lwin A, Tayebi N, LaMarca ME, Orvisky E, Sidransky E. Glucocerebrosidase mutation T369M appears to be another polymorphism. *Clin Genet* 2003; **63**: 237–238.
- Horowitz M, Pasmanik-Chor M, Ron I, Kolodny EH. The enigma of the E326K mutation in acid beta-glucocerebrosidase. *Mol Genet Metab* 2011; **104**: 35–38.
- Gan-Or Z, Amshalom I, Kilarski LL, *et al.* Differential effects of severe vs mild GBA mutations on Parkinson disease. *Neurology* 2015; **84**: 880–887.
- Zhao F, Bi L, Wang W, *et al.* Mutations of glucocerebrosidase gene and susceptibility to Parkinson's disease: an updated meta-analysis in a European population. *Neuroscience* 2016; **320**: 239–246.
- Liu G, Boot B, Locascio JJ, *et al.* Neuronopathic Gaucher's mutations accelerate cognitive decline in Parkinson's. *Ann Neurol* 2016; **80**: 674–685.
- Cilia R, Tunesi S, Marotta G, *et al.* Survival and dementia in GBA-associated Parkinson's disease: the mutation matters. *Ann Neurol* 2016; **80**: 662–673.
- Smith L, Mullin S, Schapira AHV. Insights into the structural biology of Gaucher disease. *Exp Neurol* 2017; **298**: 180–190.
- Chabas A, Gort L, Díaz-Font A, *et al.* Perinatal lethal phenotype with generalized ichthyosis in a type 2 Gaucher disease patient with the [L444P;E326K]/P182L genotype: effect of the E326K change in neonatal and classic forms of the disease. *Blood Cells Mol Dis* 2005; **35**: 253–258.
- Maor G, Cabasso O, Krivoruk O, *et al.* The contribution of mutant GBA to the development of Parkinson disease in *Drosophila*. *Hum Mol Genet* 2016; **25**: 2712–2727.
- Rolfs A, Giese A-K, Grittner U, *et al.* Glucosylsphingosine is a highly sensitive and specific biomarker for primary diagnostic and follow-up monitoring in Gaucher disease in a non-Jewish, Caucasian cohort of Gaucher disease patients. *PLoS ONE* 2013; **8**: e79732.
- Dekker N, van Dussen L, Hollak CEM, *et al.* Elevated plasma glucosylsphingosine in Gaucher disease: relation to phenotype, storage cell markers, and therapeutic response. *Blood* 2011; **118**: e118–e127.

35. Eliezer D, Kutluay E, Bussell R, Browne G. Conformational properties of alpha-synuclein in its free and lipid-associated states. *J Mol Biol* 2001; **307**: 1061–1073.
36. Hashimoto M, Hsu LJ, Xia Y, *et al*. Oxidative stress induces amyloid-like aggregate formation of NACP/ α -synuclein *in vitro*. *NeuroReport* 1999; **10**: 717.
37. Bisaglia M, Mammi S, Bubacco L. Structural insights on physiological functions and pathological effects of alpha-synuclein. *FASEB J* 2008; **23**: 329–340.
38. Spillantini MG, Schmidt ML, Lee VMY, Trojanowski JQ, Jakes R, Goedert M. $[\alpha]$ -Synuclein in Lewy bodies. *Nature* 1997; **388**: 839–840.
39. Li J-Y, Englund E, Holton JL, *et al*. Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nat Med* 2008; **14**: 501–503.
40. Masuda-Suzukake M, Nonaka T, Hosokawa M, *et al*. Prion-like spreading of pathological α -synuclein in brain. *Brain* 2013; **136**: 1128–1138.
41. Rey NL, Petit GH, Bousset L, Melki R, Brundin P. Transfer of human α -synuclein from the olfactory bulb to interconnected brain regions in mice. *Acta Neuropathol* 2013; **126**: 555–573.
42. Sacino AN, Brooks M, McGarvey NH, *et al*. Induction of CNS α -synuclein pathology by fibrillar and non-amyloidogenic recombinant α -synuclein. *Acta Neuropathol Commun* 2013; **1**: 38.
43. Luk KC, Kehm VM, Zhang B, O'Brien P, Trojanowski JQ, Lee VMY. Intracerebral inoculation of pathological α -synuclein initiates a rapidly progressive neurodegenerative α -synucleinopathy in mice. *J Exp Med* 2012; **209**: 975–986.
44. Stokholm MG, Danielsen EH, Hamilton-Dutoit SJ, Borghammer P. Pathological alpha-synuclein in gastrointestinal tissues from prodromal Parkinson's disease patients. *Ann Neurol* 2016; **79**: 940–949.
45. Shannon KM, Keshavarzian A, Mutlu E, *et al*. Alpha-synuclein in colonic submucosa in early untreated Parkinson's disease. *Mov Disord* 2011; **27**: 709–715.
46. Lebouvier T, Neunlist M, Bruley des Varannes S, *et al*. Colonic biopsies to assess the neuropathology of Parkinson's disease and its relationship with symptoms. *PLoS ONE* 2010; **5**: e12728.
47. Braak H, Tredici KD, Rüb U, de Vos RAI, Jansen Steur ENH, Braak E. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging* 2003; **24**: 197–211.
48. Dickson DW, Braak H, Duda JE, *et al*. Neuropathological assessment of Parkinson's disease: refining the diagnostic criteria. *Lancet Neurol* 2009; **8**: 1150–1157.
49. Cersosimo MG, Benarroch EE. Autonomic involvement in Parkinson's disease: pathology, pathophysiology, clinical features and possible peripheral biomarkers. *J Neurol Sci* 2012; **313**: 57–63.
50. Witt M, Bormann K, Gudziol V, *et al*. Biopsies of olfactory epithelium in patients with Parkinson's disease. *Mov Disord* 2009; **24**: 906–914.
51. Wong K, Sidransky E, Verma A, *et al*. Neuropathology provides clues to the pathophysiology of Gaucher disease. *Mol Genet Metab* 2004; **82**: 192–207.
52. Conradi NG, Sourander P, Nilsson O, Svennerholm L, Erikson A. Neuropathology of the Norrbottnian type of Gaucher disease. Morphological and biochemical studies. *Acta Neuropathol* 1984; **65**: 99–109.
53. Conradi N, Kyllerman M, Månsson JE, Percy AK, Svennerholm L. Late-infantile Gaucher disease in a child with myoclonus and bulbar signs: neuropathological and neurochemical findings. *Acta Neuropathol* 1991; **82**: 152–157.
54. Burrow TA, Sun Y, Prada CE, *et al*. Molecular genetics and metabolism. *Mol Genet Metab* 2015; **114**: 233–241.
55. Enquist IB, Bianco Lo C, Ooka A, *et al*. Murine models of acute neuronopathic Gaucher disease. *Proc Natl Acad Sci USA* 2007; **104**: 17483–17488.
56. Farfel-Becker T, Vitner EB, Futerman AH. Animal models for Gaucher disease research. *Dis Model Mech* 2011; **4**: 746–752.
57. Erikson A. Gaucher disease – Norrbottnian type (III). Neuropaediatric and neurobiological aspects of clinical patterns and treatment. *Acta Paediatr Scand Suppl* 1986; **326**: 1–42.
58. Orvisky E, Sidransky E, McKinney CE, *et al*. Glucosylsphingosine accumulation in mice and patients with type 2 Gaucher disease begins early in gestation. *Pediatr Res* 2000; **48**: 233–237.
59. Schmitz M, Alfalah M, Aerts JMFG, Naim HY, Zimmer K-P. Impaired trafficking of mutants of lysosomal glucocerebrosidase in Gaucher's disease. *Int J Biochem Cell Biol* 2005; **37**: 2310–2320.
60. Bendikov-Bar I, Maor G, Filocamo M, Horowitz M. Ambroxol as a pharmacological chaperone for mutant glucocerebrosidase. *Blood Cells Mol Dis* 2013; **50**: 141–145.
61. Ron I, Horowitz M. ER retention and degradation as the molecular basis underlying Gaucher disease heterogeneity. *Hum Mol Genet* 2005; **14**: 2387–2398.
62. Mazzulli JR, Xu Y-H, Sun Y, *et al*. Gaucher disease glucocerebrosidase and α -synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell* 2011; **146**: 37–52.
63. McNeill A, Magalhaes J, Shen C, *et al*. Ambroxol improves lysosomal biochemistry in glucocerebrosidase mutation-linked Parkinson disease cells. *Brain* 2014; **137**: 1481–1495.
64. Yap TL, Velayati A, Sidransky E, Lee JC. Membrane-bound α -synuclein interacts with glucocerebrosidase and inhibits enzyme activity. *Mol Genet Metab* 2013; **108**: 56–64.
65. Yap TL, Gruschus JM, Velayati A, Sidransky E, Lee JC. Saposin C protects glucocerebrosidase against α -synuclein inhibition. *Biochemistry* 2013; **52**: 7161–7163.
66. Shen Y-F, Tang Y, Zhang X-J, Huang K-X, Le W-D. Adaptive changes in autophagy after UPS impairment in Parkinson's disease. *Acta Pharmacol Sin* 2013; **34**: 667–673.
67. Webb JL, Ravikumar B, Atkins J, Skepper JN, Rubinstein DC. Alpha-synuclein is degraded by both autophagy and the proteasome. *J Biol Chem* 2003; **278**: 25009–25013.
68. Tan C-C, Yu J-T, Tan M-S, Jiang T, Zhu X-C, Tan L. Autophagy in aging and neurodegenerative diseases: implications for pathogenesis and therapy. *Neurobiol Aging* 2014; **35**: 941–957.
69. Rocha EM, Smith GA, Park E, *et al*. Progressive decline of glucocerebrosidase in aging and Parkinson's disease. *Ann Clin Transl Neurol* 2015; **2**: 433–438.
70. Hallett PJ, Huebner M, Brekk OR, *et al*. Glycosphingolipid levels and glucocerebrosidase activity are altered in normal aging of the mouse brain. *Neurobiol Aging* 2018; **67**: 189–200.

71. Gegg ME, Burke D, Heales SJR, *et al.* Glucocerebrosidase deficiency in substantia nigra of Parkinson disease brains. *Ann Neurol* 2012; **72**: 455–463.
72. Gegg ME, Sweet L, Wang BH, Shihabuddin LS, Sardi SP, Schapira AHV. No evidence for substrate accumulation in Parkinson brains with GBA mutations. *Mov Disord* 2015; **30**: 1085–1089.
73. Boutin M, Sun Y, Shacka JJ, Auray-Blais C. Tandem mass spectrometry multiplex analysis of glucosylceramide and galactosylceramide isoforms in brain tissues at different stages of Parkinson disease. *Anal Chem* 2016; **88**: 1856–1863.
74. Murphy KE, Halliday GM. Glucocerebrosidase deficits in sporadic Parkinson disease. *Autophagy* 2014; **10**: 1350–1351.
75. Sardi SP, Clarke J, Viel C, *et al.* Augmenting CNS glucocerebrosidase activity as a therapeutic strategy for parkinsonism and other Gaucher-related synucleinopathies. *Proc Natl Acad Sci* 2013; **110**: 3537–3542.
76. Beutler E, Gelbart T, Scott CR. Hematologically important mutations: Gaucher disease. *Blood Cells Mol Dis* 2005; **35**: 355–364.
77. Abdelwahab M, Blankenship D, Schiffmann R. Long-term follow-up and sudden unexpected death in Gaucher disease type 3 in Egypt. *Neurol Genet* 2016; **2**: e55.
78. Sidransky E, Nalls MA, Aasly JO, *et al.* Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. *N Engl J Med* 2009; **361**: 1651–1661.
79. Allen MJ, Myer BJ, Khokher AM, Rushton N, Cox TM. Pro-inflammatory cytokines and the pathogenesis of Gaucher's disease: increased release of interleukin-6 and interleukin-10. *QJM* 1997; **90**: 19–25.
80. Tantawy AAG. Cytokines in Gaucher disease: role in the pathogenesis of bone and pulmonary disease. *Egypt J Med Hum Genet* 2015; **16**: 207–213.
81. Mucci JM, Rozenfeld P. Pathogenesis of bone alterations in Gaucher disease: the role of immune system. *J Immunol Res* 2015; **2015**: 192761.
82. Pandey MK, Grabowski GA. Immunological cells and functions in Gaucher disease. *Crit Rev Oncog* 2013; **18**: 197–220.
83. Arends M, van Dussen L, Biegstraaten M, Hollak CEM. Malignancies and monoclonal gammopathy in Gaucher disease; a systematic review of the literature. *Br J Haematol* 2013; **161**: 832–842.
84. Cox TM, Rosenbloom BE, Barker RA. Gaucher disease and comorbidities: B-cell malignancy and parkinsonism. *Am J Hematol* 2015; **90**(Suppl. 1): S25–S28.
85. Mistry PK, Liu J, Yang M, *et al.* Glucocerebrosidase gene-deficient mouse recapitulates Gaucher disease displaying cellular and molecular dysregulation beyond the macrophage. *Proc Natl Acad Sci* 2010; **107**: 19473–19478.
86. Vitner EB, Farfel-Becker T, Eilam R, Biton I, Futerman AH. Contribution of brain inflammation to neuronal cell death in neuronopathic forms of Gaucher's disease. *Brain* 2012; **135**: 1724–1735.
87. Dzamko N, Geczy CL, Halliday GM. Inflammation is genetically implicated in Parkinson's disease. *Neuroscience* 2015; **302**: 89–102.
88. Kim S, Cho S-H, Kim KY, *et al.* α -Synuclein induces migration of BV-2 microglial cells by up-regulation of CD44 and MT1-MMP. *J Neurochem* 2009; **109**: 1483–1496.
89. Alvarez-Erviti L, Couch Y, Richardson J, Cooper JM, Wood MJA. Alpha-synuclein release by neurons activates the inflammatory response in a microglial cell line. *Neurosci Res* 2011; **69**: 337–342.
90. Lee E-J, Woo M-S, Moon P-G, *et al.* Alpha-synuclein activates microglia by inducing the expressions of matrix metalloproteinases and the subsequent activation of protease-activated receptor-1. *J Immunol* 2010; **185**: 615–623.
91. Park J-Y, Kim KS, Lee S-B, *et al.* On the mechanism of internalization of α -synuclein into microglia: roles of ganglioside GM1 and lipid raft. *J Neurochem* 2009; **110**: 400–411.
92. Schiess MC, Barnes JL, Ellmore TM, Poindexter BJ, Dinh K, Bick RJ. CSF from Parkinson disease patients differentially affects cultured microglia and astrocytes. *BMC Neurosci* 2010; **11**: 151.
93. Zhang W, Wang T, Pei Z, *et al.* Aggregated alpha-synuclein activates microglia: a process leading to disease progression in Parkinson's disease. *FASEB J* 2005; **19**: 533–542.
94. Gao H-M, Zhang F, Zhou H, Kam W, Wilson B, Hong J-S. Neuroinflammation and α -synuclein dysfunction potentiate each other, driving chronic progression of neurodegeneration in a mouse model of Parkinson's disease. *Environ Health Perspect* 2011; **119**: 807–814.
95. Gerhard A, Pavese N, Hotton G, *et al.* In vivo imaging of microglial activation with [¹¹C](R)-PK11195 PET in idiopathic Parkinson's disease. *Neurobiol Dis* 2006; **21**: 404–412.
96. Croisier E, Moran LB, Dexter DT, Pearce RKB, Graeber MB. Microglial inflammation in the parkinsonian substantia nigra: relationship to alpha-synuclein deposition. *J Neuroinflammation* 2005; **2**: 14.
97. Sawada M, Imamura K, Nagatsu T. Role of cytokines in inflammatory process in Parkinson's disease. *J Neural Transm Suppl* 2006; **70**: 373–381.
98. Chen H, O'Reilly EJ, Schwarzschild MA, Ascherio A. Peripheral inflammatory biomarkers and risk of Parkinson's disease. *Am J Epidemiol* 2007; **167**: 90–95.
99. Reale M, Iarlori C, Thomas A, *et al.* Brain, behavior, and immunity. *Brain Behav Immun* 2009; **23**: 55–63.
100. Gruden MA, Sewell RDE, Yanamandra K, *et al.* Immunoprotection against toxic biomarkers is retained during Parkinson's disease progression. *J Neuroimmunol* 2011; **233**: 221–227.
101. Chahine LM, Qiang J, Ashbridge E, *et al.* Clinical and biochemical differences in patients having Parkinson disease with vs without GBA mutations. *JAMA Neurol* 2013; **70**: 852.
102. Brady RO, Yang C, Zhuang Z. An innovative approach to the treatment of Gaucher disease and possibly other metabolic disorders of the brain. *J Inherit Metab Dis* 2013; **36**: 451–454.
103. Grubb JH, Vogler C, Sly WS. New strategies for enzyme replacement therapy for lysosomal storage diseases. *Rejuvenation Res* 2010; **13**: 229–236.
104. Spencer BJ, Verma IM. Targeted delivery of proteins across the blood–brain barrier. *Proc Natl Acad Sci USA* 2007; **104**: 7594–7599.
105. Sierra C, Acosta C, Chen C, *et al.* Lipid microbubbles as a vehicle for targeted drug delivery using focused ultrasound-induced blood–brain barrier opening. *J Cereb Blood Flow Metab* 2017; **37**: 1236–1250.

106. Vera M, Le S, Kan S-H, *et al.* Immune response to intrathecal enzyme replacement therapy in mucopolysaccharidosis I patients. *Pediatr Res* 2013; **74**: 712–720.
107. Shire. Study of Intrathecal Idursulfase-IT Administered in Conjunction with Elaprase® in Pediatric Patients with Hunter Syndrome and Early Cognitive Impairment – Full Text View – ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT02055118> (accessed 16/10/2018).
108. Williams IM, Wallom K-L, Smith DA, Eisa Al N, Smith C, Platt FM. Improved neuroprotection using miglustat, curcumin and ibuprofen as a triple combination therapy in Niemann–Pick disease type C1 mice. *Neurobiol Dis* 2014; **67**: 9–17.
109. Cox-Brinkman J, van Breemen MJ, van Maldegem BT, *et al.* Potential efficacy of enzyme replacement and substrate reduction therapy in three siblings with Gaucher disease type III. *J Inherit Metab Dis* 2008; **31**: 745–752.
110. Schiffmann R, Fitzgibbon EJ, Harris C, *et al.* Randomized, controlled trial of miglustat in Gaucher's disease type 3. *Ann Neurol* 2008; **64**: 514–522.
111. Zimran A, Elstein D. Management of Gaucher disease: enzyme replacement therapy. *Pediatr Endocrinol Rev* 2014; **12**(Suppl. 1): 82–87.
112. Taguchi YV, Liu J, Ruan J, *et al.* Glucosylsphingosine promotes α -synuclein pathology in mutant GBA-associated Parkinson's disease. *J Neurosci* 2017; **37**: 9617–9631.
113. Jung O, Patnaik S, Marugan J, Sidransky E, Westbroek W. Progress and potential of non-inhibitory small molecule chaperones for the treatment of Gaucher disease and its implications for Parkinson disease. *Expert Rev Proteomics* 2016; **13**: 471–479.
114. Narita A, Shire K, Itamura S, *et al.* Ambroxol chaperone therapy for neuronopathic Gaucher disease: a pilot study. *Ann Clin Transl Neurol* 2016; **3**: 200–215.